

**Overview:** This Job Aid provides step-by-step guidance for survey teams conducting schistosomiasis and STH assessment surveys in school-aged children

Step	Tasks / Requirements	Responsible Actor(s)
<b>1. Community Mobilization</b>	Introduce the survey team, explain objectives and importance of SCH and STH to the community.	District Supervisor, Program Manager, Community Health Worker (CHW), Health Workers
<b>2. Information &amp; Consent</b>	<ul style="list-style-type: none"> <li>Provide a clear explanation (purpose, procedures, risks, benefits).</li> <li>Obtain informed consent (verbal or written, as per protocol). Record refusals.</li> </ul>	Supervisor, CHW, Field Coordinators
<b>3. Eligibility Screening</b>	<ul style="list-style-type: none"> <li>Verify age (5-14 or 10-14 years).</li> <li>Confirm willingness to participate (participate (child assent obtained and parent/guardian consent from)</li> <li>Assess health status: exclude children with fever or acute illness</li> <li>Record ineligible and excluded cases with reasons</li> </ul>	Field Team
<b>4. Registration &amp; Data Capture</b>	<ul style="list-style-type: none"> <li>Assign unique participant ID</li> <li>Record demographics: name, age, sex, class level, residence/sub-district</li> <li>Record Mass Drug Administration (MDA) history if available</li> <li>Record school attendance and sanitation practices (optional questions)</li> <li>Enter data into electronic form (tablet/phone) via ESPEN Collect</li> <li>Verify data entry accuracy and completeness before submitting</li> </ul>	Data collectors
<b>5. Urine Sample Collection</b>	<ul style="list-style-type: none"> <li><b>Preparation:</b> Ensure materials ready (collection containers, labeled, gloves, disinfectant)</li> <li>Set up private collection area (latrine or designated room)</li> <li>Provide written/pictorial instructions to participants</li> <li><b>Collection:</b></li> <li>Provide ~60 mL of mid-stream urine in labeled container</li> <li>Provide stool sample (~1-2 grams or ~2-5 mL) in labeled container for STH</li> <li>Label container immediately with participant ID</li> <li>Store samples in cool location (4°C if possible)</li> </ul>	Laboratory Technician

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	<ul style="list-style-type: none"> <li>● Process within 24 hours</li> <li>● <b>Quality Control:</b> Verify all samples labeled correctly</li> <li>● Check for adequate volume</li> <li>● Record any collection issues or invalid samples</li> </ul>	
<b>6. Urine Diagnostic Testing (Hemastix or/and Urine Filtration)</b>	<p><b>Hemastix Test (if applicable):</b></p> <ul style="list-style-type: none"> <li>● Mix urine gently and perform Hemastix test per SOP to detect hematuria (indicator of S. haematobium)</li> <li>● Record result (positive/negative/invalid)</li> </ul> <p><b>Urine Filtration :</b></p> <ul style="list-style-type: none"> <li>● Use SOPs to Filter urine sample and examine microscopically for Schistosoma eggs</li> <li>● Count eggs and calculate eggs per 10 mL</li> <li>● Record result (eggs count and intensity classification)</li> </ul> <p><b>Processing:</b></p> <ul style="list-style-type: none"> <li>● Process samples same day or store appropriately</li> <li>● Record all results on urine form</li> </ul>	<p>Laboratory Technician</p>
<b>7. Stool Diagnostic Testing (Kato-Katz Method)</b>	<ul style="list-style-type: none"> <li>● <b>Preparation:</b> Prepare thick smears using Kato-Katz template (41.7 mg template standard)</li> <li>● Samples should be fresh or stored appropriately</li> <li>● <b>Processing:</b> Apply stool to template on microscope slide</li> <li>● Press cellophane (pre-soaked in glycerol) over stool</li> <li>● Allow to clear for 24 hours</li> <li>● Examine microscopically at 40x and 100x magnification</li> <li>● <b>Reading &amp; Quantification:</b> Count Schistosoma mansoni and other helminth eggs</li> <li>● Multiply by 24 to calculate eggs per gram (epg)</li> <li>● Classify intensity: Light (&lt;100 epg), Moderate (100-399 epg), Heavy (≥400 epg)</li> <li>● Record result on Kato-Katz form</li> <li>● <b>Storage:</b> Store prepared slides for quality control re-reading</li> </ul>	<p>Laboratory Technician</p>
<b>8. Data Verification</b>	<ul style="list-style-type: none"> <li>● Cross-check test results for completeness and accuracy</li> <li>● Verify all participant IDs match across forms</li> <li>● Check for missing data or anomalies</li> <li>● Daily data review and cleaning</li> <li>● Secure backup of electronic data</li> <li>● Record GPS coordinates of survey sites</li> <li>● Verify survey dates and team members</li> </ul>	<p>Data Collectors, Supervisor, Data Manager</p>

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<b>9. Quality Control &amp; Re-reading</b>	<ul style="list-style-type: none"> <li>● Re-read ≥10% of all negative urine slides and 100% of positive slides</li> <li>● Re-read ≥10% of all negative Kato-Katz slides and 100% of positive slides</li> <li>● Investigate discrepancies between initial and re-read results</li> <li>● Cross-check all labels and participant IDs</li> <li>● Document quality control findings</li> </ul>	Supervisor, Senior Lab Technician
<b>10. Waste Disposal &amp; Safety</b>	<ul style="list-style-type: none"> <li>● Collect sharps (lancets, needles) in safety boxes</li> <li>● Place all biological waste in biohazard bags</li> <li>● Follow needlestick injury protocol if exposure occurs</li> <li>● Maintain hand hygiene throughout survey</li> <li>● Use appropriate PPE (gloves, aprons) at all times</li> <li>● Reinforce universal precautions at all times.</li> <li>● Document any accidents or injuries</li> </ul>	All Team Members
<b>11. End of Process</b>	<ul style="list-style-type: none"> <li>● Thank all participants</li> <li>● Provide health education: SCH transmission, prevention, importance of treatment</li> <li>● Identify infected individuals and inform of treatment availability through MDA program</li> <li>● Maintain confidentiality of individual results while sharing aggregate findings</li> </ul>	Field Team, Health Workers